



ELISA TEST | In vitro analysis

for the quantitative detection of T-2 in grains, cereals and animal feed

ProGnosis Biotech S.A. is ISO 9001:2015 certified by TÜV Hellas (TÜV NORD).

Use only the current version of Product Data Sheet enclosed with the kit.

Bio-Shield T-2, B4148/B4196, is an immunoassay method that determines the T-2 in grains, cereals and other commodities including animal feed. The ELISA kit contains all reagents required for the immunoassay method. The ELISA test is adequate for 48/96 definitions (standards are included). A spectrophotometer for microtiter ELISA plate is required.

Matrices:

Cereals: Alfalfa, Barley, Biscuits, Bran sticks, Brown rice, Corn, Corn flakes, Corn flour, Corn gluten, Corn gluten meal, Cottonseed, DDGS, Dried sunflower, Oat Bran, Oat flakes, Oats, Pet food, Popcorn, Raw rye, Rice, Rye flour, Sesame, Sorghum, Soy beans, Soy flour, Wheat, Wheat bran, Wheat flour

Other: Beans, Cocoa, Milk.

- Sample preparation: extraction
- Test time (incubation time after samples and reagents preparation): 20min
- Standard curve range: 0 - 500ppb
- Shelf life: 12 months
- Storage: 2-8°C

Specifications

- The LOD of the method is 3.3 ppb T-2.
- The LOQ of the method is 6.0 ppb T-2.
- The recovery of spiked extractions-matrices was 96.1% (CV = 4.1%).
- IC50 = 40 - 160 ppb
- Each standards duplicates mean CV ≤ 6%
- The cross-reaction of the anti-T-2 antibody with T-2, HT-2, T-2 Triol and T-2 Tetraol is 100, <0.1%, <0.1% and <0.1% respectively.

1. Description

Bio-Shield T-2 is an ELISA test for the detection of Trichothecene T-2 in grains, cereals and animal feed.

2. General Information

T-2 toxin belongs to the group of trichothecenes. This group of mycotoxins are produced mainly by fungi of the genus Fusarium which is toxic to humans and animals. Agricultural commodities are frequently infected by this fungus. It is frequently implicated in cytotoxic and immunosuppressive disorders of farm animals and occasionally in pathogenetic syndromes in humans. Both in man and in animals T-2 toxin can cause alimentary toxic aleukia. Most controlling government agencies worldwide have regulations or recommendations regarding the amount of T-2 allowable in human and animal foodstuffs. Accurate and rapid determination of T-2 presence in commodities is of paramount importance.

3. Principle of the Method

The quantitative test is based on the enzyme linked immunosorbent assay principles. The wells of the microtiter strips are coated with T-2 specific antibodies. Toxins are extracted from a ground sample with methanol 70%. Trichothecene standards or samples and T-2-HRP conjugate (detection solution) are added into the coated wells. T-2-HRP conjugate binds to the binding sites of coated antibodies that are not already occupied by T-2 of standards or samples. Any unbound T-2-HRP conjugate of detection solution is removed in a washing step. A chromogen substrate is added to the wells resulting in the progressive development of a blue colored complex with the detection antibody. The color development is then stopped by the addition of acid turning the resultant final product yellow. The measurement is made photometrically at 450 nm and the intensity of the produced colored complex is indirectly proportional to the concentration of T-2 present in the samples and standards.

4. Reagents Provided

Bio-Shield T-2 ELISA kit contains sufficient reagents and materials for 48/96 measurements (including standard tests).

Reagents (Store at 2-8°C)	Quantity for 48 wells	Quantity for 96 wells	State	Vial cap color
Single-Break Strip Plate	48 wells	96 wells	Ready to use (precoated)	-
Dilution Microwells	48 wells	96 wells	Ready to use (green color)	-
Sealing film	2 sheets	2 sheets	Ready to use	-
Standards 1-5 (0, 2, 8, 20 and 50ppb of T-2 in organic solution) (correspond to 0, 20, 80, 200 and 500ppb)	5 plastic vials (each 1.5ml)	5 plastic vials (each 1.5ml)	Ready to use	Brown
T-2 Detection Solution	1 plastic vial (12ml)	2 plastic vials (each 12ml)	Ready to use	Green
Wash Buffer	1 plastic vial (50ml)	1 plastic vial (50ml)	20X Concentrate (dilute in distilled water)	White
TMB Substrate	1 plastic vial (6ml)	1 plastic vial (12ml)	Ready to use	Brown
Stop Solution	1 plastic vial (6ml)	1 plastic vial (12ml)	Ready to use	White

5. Materials required but not provided

- A grinder sufficient to render sample to particle size of fine instant coffee
- Balance with 0 - 50 g measuring capability and Graduated cylinder - 100 mL
- Methanol (70 mL reagent grade per sample) and Distilled or deionized water
- Filter Paper Whatman #1 or equivalent, Filter Funnel and Miscellaneous laboratory plastic or glass tubes 50 - 125 mL

- Vortex mixer and Microtiter plate reader fi ted with 450 nm filter
- 100, 200 and 1000 µL adjustable single channel micropipettes with disposable tips (a repetitive pipette of 100µL is acceptable for the steps of TMB and Stop Solution)
- 50 - 300 µL multi-channel micropipette with disposable tips and reservoirs

6. Storage Instructions

Store kit reagents between 2 and 8°C (35 - 46°F). Do not freeze any components provided. Reseal immediately the unused strips of the microtiter plate in the bag together **with the desiccant bag** provided and store at 2 - 8°C. After use remaining reagents should be returned to cold storage (2 - 8° C). Expiry of the kit and reagents is stated on the labels respectively and no quality guarantee is accepted after the expiration date. The expiry of the kit components can only be guaranteed if the components are stored properly as well as if the reagent is not contaminated by the first handling, in case of repeated use of one component. Because of the colorless TMB Substrate and standards 1-7 light sensitivity, avoid the exposure to direct light. Do not interchange individual reagents between kits of different lot numbers.

7. Safety and Precautions for use

- Avoid any skin contact with T-2 standards, Stop Solution (8% H₃PO₄) and TMB (toxic). **Use gloves.** In case of contact, wash thoroughly with water.
- All reagents should be warmed in room temperature before use and covered when not in use. **Use a clean disposable plastic pipette tip for each reagent, in order to avoid cross contamination. When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells.**
- Use a clean plastic container to prepare the wash buffer and all residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper into the well. Read the absorbance within 60 minutes after completion of the assay.

8. Indication of corruption of kit reagents

- The bluish coloration of the chromogen substrate before the ELISA test.
- A value of less than 0.7 absorbance units (ABS 450nm) for the Standard 1 (St1).

9. Sample and reagents preparation

9.1 Reagents preparation

- Prepare the Extraction Solution (70% Methanol) by adding 30ml of distilled or deionized water to 70ml of methanol (reagent grade) for each sample to be tested.
- Dilute the 20X solution concentrate 20 fold with distilled water to give a **1X** working solution.

9.2 Samples preparation

Preparation of Wash Buffer 1X: In case of the occurrence of crystals in the Wash Buffer, the warming by gentle dismantling (using hands) of the crystals is needed. Pour entire content of the solution concentrate (50ml) into a clean 1000ml graduated cylinder, rinse the vial with distilled or deionized water and pour the content again into the cylinder and fill to a final volume of 1000ml with distilled or deionized water. Mix gently to avoid foaming, transferring the final solution from cylinder to a clean bottle and back two times. The clean bottle with **1X Wash Buffer** working solution can be left out of the refrigerator during the method procedure and subsequent be stored 2 - 8°C for one month.

9.2.1 Ground samples

1. The sample must be collected according to established sampling techniques. Grind a representative sample to the particle size of fine instant coffee (50% passes through a 20 mesh screen).
 2. Weigh out a 20 g ground portion of the sample and add 100 mL of the Extraction Solvent (70% methanol) and mix in a blender for a minimum of 2 minutes. **The ratio of sample to extraction solvent is 1:5 (w/v).**
 3. Allow the particulate matter to settle, filter 5 - 10 mL of the extract through a Whatman #1 filter paper (or equivalent), collect the filtrate and dilute 2 times with deionized water (example: 1 mL filtrate + 1 mL deionized water). **The ratio of sample to 35% methanol is 1:10 (w/v).**
 4. Use 100 µL of each final diluted filtrate directly in the immunoassay.
- NOTE 1:** The extracted sample should have pH value of 6.2 - 7.5. If the pH is less than 6.2, it should be neutralized using NaOH.
- NOTE 2:** In case the user makes an additional dilution 1:1 of filtrate with 70% methanol (example: 500 µL filtrate + 500 µL 70% methanol) the range of quantification becomes 0 - 1000ppb. Then, dilute this 2-fold diluted filtrate 2 times with deionized water (example: 1 mL 2-fold diluted filtrate + 1 mL deionized water) and use 100µl directly in the immunoassay. Multiply the final T-2 ppb result x 2.

9.2.2 Milk Samples

After centrifugation at 3000xg at 4°C for 10 min, remove the upper fat layer, dilute the defatted milk sample 5 times with 35% methanol (**1ml of milk + 4ml of 35% methanol**) and vortex. Use 100µl of each diluted milk sample directly in the immunoassay and divide the final T-2 ppm result by 2. The LOD is 1.7ppb and the LOQ is 3ppb.

10. Method Procedure

10.1 Assay Design: Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for standards. Considering that each sample and standard can be tested in single or in duplicate, create a layout.

NOTE: Do not use more than 48 wells (six strips) in a single experiment.

CAUTION: Use the standards positions as the **Example plate** layout below **NECESSARY** and note positions of samples that can be set to all remaining empty wells of layout.

	1	2	3	4	5	6	7	8	9	10	11	12
A	St1	St1										
B	St2	St2										
C	St3	St3										
D	St4	St4										
E	St5	St5										
F												
G												
H												

Example plate layout (example for a 5 point standard curve)

10.2 Bring all reagents to room temperature (19 - 24°C) before use. Remove the standards (**Standard 1-5**) and place two Dilution Microwells (green) in a microwell holder for each Standard and Sample to be tested in duplicate. Place an equal number of Antibody Coated Microtiter Wells in another microwell holder. Immediately reseal the unused strips of the microtiter plate in the bag together with the desiccant bag provided. The samples should be stored in a cool place.

10.3 Add **200µl** of **T-2 Detection Solution** to each Dilution Well.

10.4 Using new pipette tip for each, add **100µl** of each Standard (**Standard 1 - 5**) and prepared sample in duplicate (see Chapter 9) to appropriate Dilution Well containing the **T-2 Detection Solution**. Mix by priming pipetting at least 5 times.

10.5 Using a multichannel pipette, transfer **100µl** of contents from each Dilution Microwell to a corresponding Antibody Coated Microtiter Well. Incubate at room temperature for **10 minutes**.

10.6 Remove the sealing film and wash the plate as follows: Aspirate the liquid from each well into the sink and tap the holder of microwells upside down strongly (four times in a row) on an absorbent paper to insure the complete removal of liquid from the wells. Dispense **300µl** of **Wash Buffer 1X** (see 9.1) into each well with wash bottle or multichannel micropipette using the proper reagent reservoir and shaking the plate manually for a few seconds. Repeat this process for another three times (**total 4 times**). **CAUTION:** It is important to not allow microwells to dry between working steps.

10.7 Aspirate the liquid as described above and add **100µl** per well of **TMB Substrate** (pour 1ml per 8 wells in a reservoir). Cover the microwells with the sealing film, shaking the plate manually for a few seconds and incubate in the dark at room temperature for **10 minutes**.

10.8 Remove the sealing film and add **100µl** per well of the **Stop Solution** to each well (pour 1ml per 8 wells in a reservoir). Mix gently by shaking again the plate manually.

10.9 Measure the absorbance at 450nm. Read the absorbance value of each well (within 60 minutes after the step 10.8) on a spectrophotometer using 450 nm as the primary wavelength and optionally 620nm as the reference wave length (610nm to 650nm is acceptable).

11. Data Analysis

• Automatically

An assigned software, the **Prognosis-Data-Reader**, is available for free (contact: info@prognosis-biotech.com) download in order to evaluate the Bio-Shield T-2 ELISA kit. The evaluation is carried out by a simple transfer of data values after the measurement.

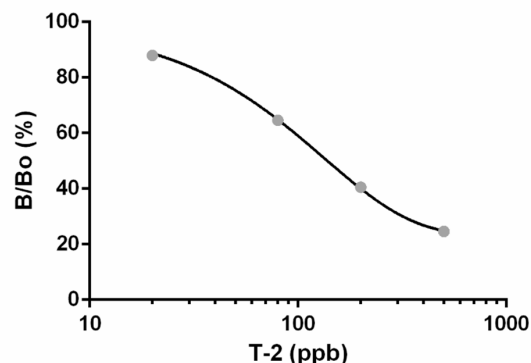
• Manually

Calculate the average absorbance values for each set of duplicate standards and samples. Ideally duplicates should be within 10% of the mean. Use the following calculation:

$$\frac{\text{Standard or sample absorbance}}{\text{Standard 1 absorbance}} \times 100 = \% \text{ Binding}$$

The standard 1 is equal to 100% and the absorbance values are quoted in percentages. The concentration of Trichothecene (ppb) in each sample is determined by extrapolating OD values against concentrations of Trichothecene in standard solutions using a two phase exponential decay standard curve with logarithmic X axis.

12. Example of Standard Curve (0 - 500ppb)



13. Performance Evaluation

13.1 Reference Materials

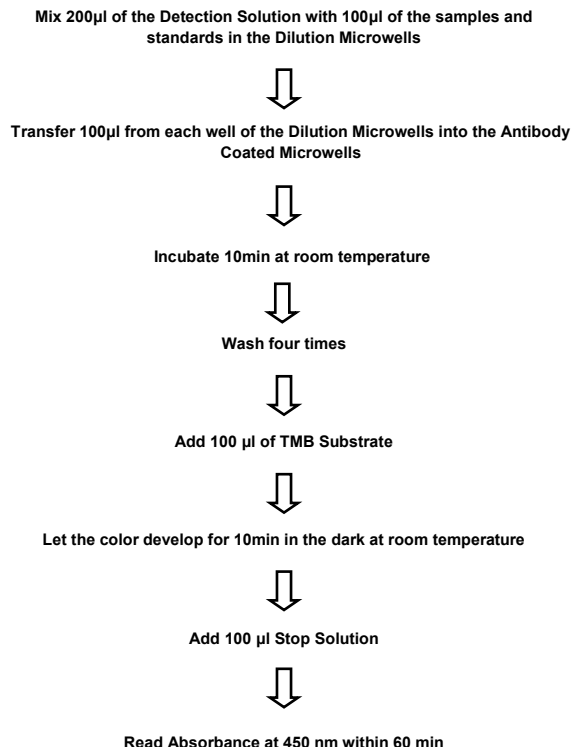
Several reference materials are being used for the evaluation of each product of Prognosis Biotech S.A. in the context of Quality Control performed by Quality Control Department. Please request a validation report, including the results, at info@prognosis-biotech.com.

13.2 Proficiency Tests

All products participate frequently in Proficiency Tests. For more information, visit the individual product page in our website: www.prognosis-biotech.com

14. Method Summary

Total procedure time (after samples and reagents preparation): 20min.



VERSION N3

CAT.NUMBER: B4148/B4196

STORAGE: 2-8°C



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Other: Beans, Cocoa, Milk.

- Sample preparation: extraction
- Test time (incubation time after samples and reagents preparation): 20min
- Standard curve range: 0 - 500ppb
- Shelf life: 12 months
- Storage: 2-8°C

All immune assays supplied by ProGnosis Biotech S.A., are warranted to meet or exceed our published specification when used under normal conditions in your laboratory. If the product fails during the stated period, a replacement product will be issued.

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