

RAPID TEST ADENOVIRUS

Lateral Flow Test | IVD

For the detection of Adenovirus antigen in human nasal or nasopharyngeal specimen.

This Lateral Flow test kit is manufactured by ProGnosis Biotech S.A. and complies with the specifications on the Standard EN ISO 13485:2016

Use only the current version of Product Data Sheet enclosed with the kit.

Rapid Test Adenovirus, V18XX, is a qualitative Lateral Flow test for the detection of Adenovirus antigens in nasal or nasopharyngeal specimen.

The Lateral flow kit contains all reagents required for the immunoassay method.

Specimen: Nasal or nasopharyngeal swab.

- For professional use only

- For in vitro diagnostic use only
- Rapid Test Adenovirus is a sensitive screening assay for the detection of Adenovirus antigen. Results should not be used as the only source to diagnose or to determine infection status.
- Negative result do not rule out Adenovirus infection
- Test should only be conducted by medical personnel
- Test time (incubation time after specimens preparation): 15 min
- Shelf life: 2 years
- Storage: 4-30°C

1. Description

Rapid Test Adenovirus is a qualitative, lateral flow immunoassay designed to detect the presence or absence of Adenovirus antigens in nasal or nasopharyngeal swab specimens directly collected.

2. Principal of the assay

In this test, antibodies specific to Adenovirus are coated on the test line region (Test line) of the nitrocellulose membrane. During testing, antigens in the specimen react with the antibodies that are coated onto gold nanoparticles. The mixture migrates up the membrane to react with the antibodies immobilized on the membrane and generate a colored line in the test region T. The presence of the colored Test line indicates a positive result. To serve as a procedural control, a colored line will always appear in the control region if the test has been performed properly.

3. Introduction

Adenoviruses are common in humans, usually cause respiratory illness but they are also responsible for gastroenteritis, conjunctivitis, cystitis. Symptoms of respiratory illness range from the common cold syndrome to pneumonia. Adenovirus does not show seasonal infection, as they appear anytime through the year and in all ages. In children adenovirus causes infections of the upper respiratory system, gastroenteritis and conjunctivitis. The virus can spread through droplets from a cough, talk or sneeze.

4. Reagents Provided

	V1801	V1805	V1820
Reaction device	1	5	20
Prefilled extraction tube with Running Buffer	1	5	20
Sterile swabs	1	5	20
Carton case	-	-	1
Instruction of use	1	1	1

5. Materials required but not provided

- Clock or Timer
- Gloves and face protection shield
- Container for biohazardous waste

6. Storage Instructions

Store kit components between 4 and 30°C (39.2 - 96°F). Do not freeze any components provided. Expiry of the kit and reagents is stated on their labels and no quality guarantee is accepted after the expiration date. The expiry of the kit components can only be guaranteed if the components are stored properly and the reagent is not contaminated prior handling.

7. Safety and Precautions for use

7.1 Health and safety precautions

- Use gloves, protective clothing and eye/face protection and handle appropriately with the requisite Good Laboratory Practices. The product must only be used by qualified personnel.
- **WASTE MANAGEMENT:** Dispose of all specimens and materials used to perform the test as bio-hazard waste. Laboratory chemical and biohazard wastes must be handled and discarded in accordance with all local, state, and national regulations.
- During the specimen collection human source material spills should be also treated as potentially infectious. Spills should be immediately decontaminated, including the spill area, materials and any contaminated surfaces, with an appropriate chemical disinfectant and should be wiped away.
- Do not re-use any of the kit components as they are single-use only.
- Sterile swabs must be used only for nasal or nasopharyngeal specimen. Avoid to touch the tip of the swab.
- Do not eat, drink or smoke in the area where the specimens and the kit are stored and handled.
- All positive results should be processed following local news and regulations.

7.2 Precautions related to the procedure

- In accordance with Article 1, Paragraph 2b of European Directive 98/79/EC, the use of in-vitro diagnostic medical devices is envisaged by the manufacturer to ensure the suitability, performance, and safety of these products. Consequently, the testing procedure, information, precautions, and warnings in the instructions for use must be followed rigorously. No changes to the test procedure are permitted, nor is any use in combination with other products not approved by the manufacturer. The user is solely responsible for any such changes. The manufacturer is not responsible for false results nor incidents arising as a result of these.
- Do not use the kit if the packaging of components is damaged, if there is an expired reagent or if the desiccant bag is absent inside the vial containing the sticks.
- All reagents should be warmed in room temperature before use.
- Cover or cap all reagents when not in use.
- Do not mix and interchange different specimens.
- Do not interchange individual reagents between kits of different lot numbers.

8. Specimen collection

8.1 Nasal Mid-Turbinate specimen collection

Tilt the patient's head back 70 degrees. Remove a sterile swab from the pouch. While gently rotating it, insert the swab less than one inch (about 2 cm) into patient's nostril (until resistance is met at the turbinates). Rotate the swab five times against the nasal wall then slowly remove from the nostril. Using the same swab repeat the collection procedure with the second nostril. (Figure 1.)

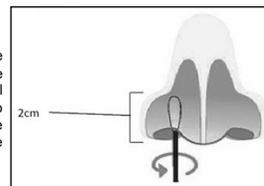


Figure 1. Nasal swab procedure

8.2 Nasopharyngeal specimen collection

Tilt the patient's head back 70 degrees. Remove a sterile swab from the pouch. Place the swab into one of the patient's nostrils. When it reaches the posterior nasopharynx rotate 3 to 5 times over the surface and then remove it slowly while rotating it (Figure 2.)

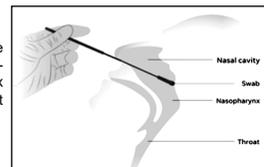


Figure 2. Nasopharyngeal swab procedure

9. Method Procedure

- 9.1 Calculate the number of swabbing sticks and tubes needed, according to the number of samples to collect.
 - 9.2 After the specimen collection (see Chapter 8), place the swab in the extraction tube, rotate the swab forcefully against the side of the tube for 1min. Best results are obtained when the specimen is vigorously extracted in the solution.
 - 9.3 Remove the swab, squeezing the sides of the tube to extract as much liquid as possible.
 - 9.4 Discard the swab.
 - 9.5 Close the extraction tube with the dropper cup. Add 3 drops in the circular window of the cassette.
 - 9.6 After 15 minutes, the test stick can be visually read and interpreted according to the corresponding figure.
- Note:** The test result should not be read and interpreted after 30 minutes.

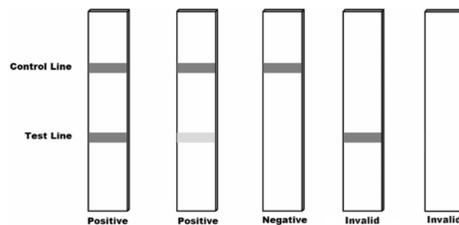
10. Interpretation of results

Note: For internal procedure purposes two colored lines are present on the result window of the Rapid Test Adenovirus. The colored lines have no effect on the product's performance since they are washed away during the experiment.

Positive: Two visible colored bands appear at both Test line (T) and Control (C) line. It indicates a positive result for the Adenovirus antigen in the specimen.

Negative: One visible colored band appears at Control line. It indicates that the concentration of the antigen is zero or below the detection limit of the test.

Invalid: No colored band appears at Control line no matter whether it appears at Test line or not.



Interpretation of results

11. Limitations

- The test procedure, precautions and interpretation of results for this test must be followed strictly when testing.
- After specimen collection the swab must be extracted as soon as possible. Otherwise they may be stored dry in their original packaging for up to 2 hours at RT. When using a viral transport medium, extract AGAIN the swab in the extraction tube that contains Prognosis Running Buffer. When using the transport media, the test sensitivity can be reduced due to excessive dilution of specimen.
Note: the transport medium should not contain chaotropic substances such as Guanidinium thiocyanate.
- After the extraction specimens should be tested as soon as possible. Otherwise they can be stored at room temperature 20-25°C (68-77°F) for two hours.
- The test should be used for the detection of Adenovirus antigen ONLY in nasal or nasopharyngeal swab specimens.
- Failure to follow the guidelines for proper specimen collection, test procedure and interpretation of test results may adversely affect test performance and/or produce invalid result.

- USE ONLY the sterile swabs that are provided in the kit for the specimen collection.
- During specimen collection avoid contact with bleeding areas and excess of mucus as both of them may give a false positive result due to interference with the test performance.
- Positive results indicate the presence of Adenovirus antigens but a diagnosis of an infection should only be made by a physician evaluating all clinical and laboratory findings and must be based in the correlation of the results with further clinical observations.
- A negative test result may occur if the level of extracted antigen in a specimen is below the sensitivity of the test or if a poor quality specimen is obtained.
- Positive test results do not rule out co-infection with other pathogens.
- The Rapid Test Adenovirus will indicate the presence of Adenovirus in the specimen from both viable and non-viable virus.

12. Immunoassay Performance

12.1 Cross-reactivity

In order to determine the cross reactivity of Rapid Test Adenovirus, an evaluation was performed; no cross reactivity against organism, pathogens that could cause infections was observed.

Microorganism	Concentration	Result
Alpha coronavirus 229E	4.68 x 10 ⁴ TCID ₅₀ /mL	Negative
Alpha coronavirus NL63	1.70 x 10 ⁵ TCID ₅₀ /mL	Negative
Beta coronavirus OC43	5.01 x 10 ⁵ TCID ₅₀ /mL	Negative
Escherichia Coli O157	6.4x10 ⁶ CFU/ml	Negative
Influenza A virus	1.51 x 10 ⁶ TCID ₅₀ /mL	Negative
Influenza B virus	5.01 x 10 ⁵ TCID ₅₀ /mL	Negative
Listeria monocytogenes	2.5 x10 ⁶ CFU/ml	Negative
Respiratory Syncytial Virus type A	4.68 x 10 ⁴ TCID ₅₀ /mL	Negative
Respiratory Syncytial Virus type B	1.17 x 10 ⁶ TCID ₅₀ /mL	Negative
Salmonella enteritidis	3.6x10 ⁶ CFU/ml	Negative
SARS-Cov-2	1.15 x 10 ⁷ TCID ₅₀ /mL	Negative
Streptococcus pneumococcal	4.2x10 ⁶ CFU/ml	Negative
Streptococcus pyogenes	3.6x10 ⁶ CFU/ml	Negative

12.2 Interference Data

The following substances showed no significant interference on the test results of Rapid Test Adenovirus.

No	Interfering Substances	Final Test
1	Azithromycin	84 mg/ml
2	Amoxicillin	54 mg/L
3	Albuterol	0.05 mg/L
4	Acarbose	0.3 mg/L
5	Chlorpheniramine	0.8 mg/L
6	Chlorothiazide	27 mg/L
7	Rheumatoid factor	200 IU/ml
8	Triglycerides	1.5 mg/L
9	Hemoglobin	100 mg/L
10	Human Chorionic Gonadotropin Hormone (pregnancy)	10-fold dilution
11	Ibuprofen	219 mg/L
12	Xylometazoline (Otriven)	10%
13	Acetylsalicylic Acid	3 mg/ml
14	Mucin	0.5%

12.3 High Dose Hook Effect

No high dose hook effect was observed up to 1.02 x 10⁸ TCID₅₀/mL of heat inactivated Adenovirus type 7A and up to 2.57 x 10⁶ TCID₅₀/mL of heat inactivated Adenovirus type 1, with the Rapid Test Adenovirus.

12.4 Limit of Detection

The lowest detectable concentration of an analyte in a method is known as LOD. In this case, serial dilution of heat inactivated viruses were used in order to determine the limit of detection in Rapid Test Adenovirus. The LOD is the level at which 95% of the replicates are characterized as positive. The results of 20 replicates are shown below.

LOD for Adenovirus type 7A : 5.1×10^3 TCID₅₀/mL

LOD for Adenovirus type 1 : 2.57×10^3 TCID₅₀/mL

Adenovirus type 7A

Concentration (TCID ₅₀ /mL)	Positive Replicates	Visual interpretation of results
1.02×10^8	20 / 20	Strong positive
1.02×10^7	20 / 20	Strong positive
1.02×10^6	20 / 20	Strong positive
1.02×10^5	20 / 20	Strong positive
1.02×10^4	20 / 20	Positive
5.1×10^3	20 / 20	Positive
25.5×10^2	7 / 20	Negative

Adenovirus type 1

Concentration (TCID ₅₀ /mL)	Positive Replicates	Visual interpretation of results
2.57×10^8	20 / 20	Strong positive
2.57×10^7	20 / 20	Strong positive
2.57×10^6	20 / 20	Strong positive
2.57×10^5	20 / 20	Positive
2.57×10^4	20 / 20	Positive
2.57×10^3	20 / 20	Positive
1.28×10^3	4 / 20	Negative

12.5 Clinical performance characteristics

In order to determine the clinical performance of the Rapid Test ADENOVIRUS, a total amount of 690 specimens both with nasopharyngeal and nasal specimen collection were tested.

12.5.1. Nasopharyngeal specimens

659 negative and 31 positive specimens (total amount 690) confirmed with RT-PCR assay [ADENOVIRUS R-GENE(Biomerieux)] were tested. The results are presented below.

Rapid Test Adenovirus	Real-time PCR		
	Positive	Negative	Total
Positive	28	5	33
Negative	3	654	657
Total	31	659	690

Clinical Diagnostic Specificity: 99.2%

Clinical Diagnostic Sensitivity: 90.3%

12.5.2. Nasal specimens

659 negative and 31 positive specimens (total amount 690) confirmed with RT-PCR assay [ADENOVIRUS R-GENE(Biomerieux)] were tested. The results are presented below.

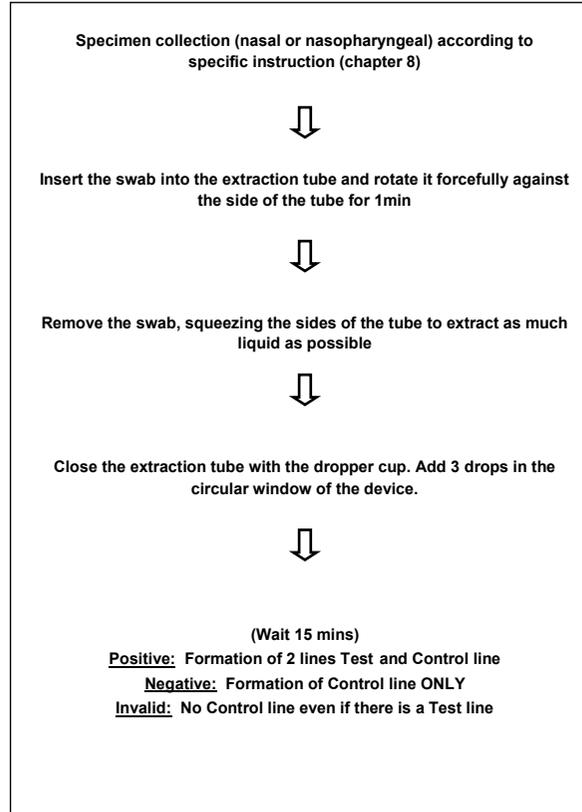
Rapid Test Adenovirus	Real-time PCR		
	Positive	Negative	Total
Positive	27	5	33
Negative	4	654	657
Total	31	659	690

Clinical Diagnostic Specificity: 99.24%

Clinical Diagnostic Sensitivity: 87.10%

13. Method Summary

Total procedure time (after specimens preparation): 15 min.



REF V18XX



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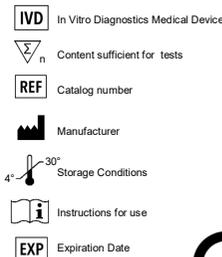
Specimen: Nasal or nasopharyngeal swab.

The extracted specimen can be used with the kit codes V15XX, V16XX, V17XX, V18XX

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- Storage: 4-30°C

14. References

- Centers for Disease Control and Prevention. <https://www.cdc.gov/adenovirus/outbreaks.html>
- <https://www.cdc.gov/flu/symptoms/flu-vs-covid19.htm>.
- MARCELA ECHAVARRIA, JOSE L. SANCHEZ, et al. "Rapid Detection of Adenovirus in Throat Swab Specimens by PCR during Respiratory Disease Outbreaks among Military Recruits", Journal of Clinical Microbiology, Feb. 2003, Vol. 41, No. 2, p. 810-812
- Harrington et al. Rapid Detection and Identification of Human Adenovirus Species by Adenoplex, a Multiplex PCR-Enzyme Hybridization Assay doi: 10.1128/JCM.42.9.4072-4076.2004



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